



ISSN: 0976-3376

Available Online at <http://www.journalajst.com>

ASIAN JOURNAL OF
SCIENCE AND TECHNOLOGY

Asian Journal of Science and Technology
Vol. 6, Issue 04, pp. 1316-1318, April, 2015

RESEARCH ARTICLE

SUB ACUTE DERMAL TOXICITY OF METALAXYL WITH SPECIAL REFERENCE TO OXIDATIVE STRESS IN WISTAR RATS

¹Mudasir sultana, ²Shahid Prawez, ^{3*}Muneer Ahmad Dar, ⁴Mahrukh Ahmad and ⁵Sania Naseem

^{1, 4, 5}Division of Veterinary Pharmacology and Toxicology, FVSc and AH, SKUAST-Jammu

²Division of Veterinary Pharmacology and Toxicology, BHU

³Department of Veterinary Pharmacology and Toxicology, MJF College of Veterinary and Animal Sciences, Chomu, Jaipur

ARTICLE INFO

Article History:

Received 29th January, 2015

Received in revised form

02nd February, 2015

Accepted 15th March, 2015

Published online 30th April, 2015

Key words:

Metalaxyl,

Dermal,

Sub acute

Oxidative stress.

ABSTRACT

The present study was aimed to evaluate the oxidative stress potential of metalaxyl after its dermal application for a period of 30 days in wistar rats. Rats were divided into two groups with six rats in each group. Group I served as control and were dermally applied with distilled water. Group II received dermal treatment of metalaxyl @ 350mg/Kg.bwt (1/10th LD₅₀) after dissolution in water. Significant increase in Lipid peroxidation was observed on 30th day of treatment in metalaxyl treated group as compared to control. Blood glutathione decreased significantly in metalaxyl treated animals as compared to control ones. There was significant decrease in the activities of SOD, CAT and GPx on 30th day in metalaxyl treated animals as compared to control group. However, no significant change was observed in GST activity which depicted decreasing trend as compared to control. The present study follows that metalaxyl on dermal application can produce inevitable changes in the form of oxidative stress even for short period of 30 days and therefore warns about injudicious use of the pesticide.

Copyright © 2015 Mudasir Sultana et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

INTRODUCTION

Fungicides are dangerous category of pesticides and are sprayed as many as twenty times in a year on fresh fruits and vegetables during humid conditions to prevent growth of mold and fungus affecting productivity (Science News, 1994). Every organ system in the human body is vulnerable to the toxic effects of these synthetic chemicals. It is well understood that the indiscriminate use of agrochemicals under conventional agriculture not only causes severe health hazards for human beings but also has numerous other side effects on the environment including destruction of the biodiversity. Therefore, animal toxicity studies are most important part in the assessment of safety of these chemicals. Metalaxyl, a systemic benzenoid fungicide belongs to chemical group of acylalanine having IUPAC name methyl *N*-(2-methoxyacetyl)-*N*-(2,6-xylyl)-DL-alaninate, with molecular formula C₁₅H₂₁NO₄ (FAO, 1995). Metalaxyl is used to control soil-borne diseases caused by *Phytophthora* and *Pythium* on fruits, cotton, soyabean, peanuts, ornamentals and grasses (Sukul and Spittler, 2000). Few animal studies suggest that the metalaxyl produces hepatotoxicity and various hazardous effects in

mammalian animals when given orally for longer period of time (Howard, 1991 and Okdah, 2005). However, there is dearth of studies to evaluate the dermal toxic potential of this commonly used fungicide for different time intervals. Therefore, present study was an attempt to appraise the toxic potential of this chemical after its dermal application for a period of 30 days in wistar rats.

MATERIALS AND METHODS

Experimental animals

The study on effects of metalaxyl was conducted on healthy wistar rats of either sex weighing 200 to 250 g procured from Indian Institute of Integrative Medicine, CSIR Lab, Jammu. The animals were provided standard pelleted ration and clean drinking water *ad libitum*. All the animals were maintained under standard managemental conditions. A daily cycle of 12 h of light and 12 h of darkness was provided to animals. Prior to start of experiment, the rats were acclimatized in the laboratory conditions for a period of more than 3 weeks. All the experimental animals were kept under constant observation during entire period of study. The experiment was conducted strictly in accordance to the Institutional Animal Ethics committee.

*Corresponding author: Abrar Ahmed

Division of Veterinary Pharmacology and Toxicology, MJF College of Veterinary and Animal Sciences, Chomu, Rajasthan.

Insecticide used

Metalaxyl (35%WS) was commercially obtained from Jai Shree Rasayan Udyog Limited, Delhi as Srilaxyl 35 in 100 gram pack. The metalaxyl was applied dermally @ 350 mg/kg (1/10th LD₅₀) (US, HSD, 1995) on interscapular region as per method described by Punareewattana *et al.* (2001)

Experimental design and dosage

Rats of either sex were divided into two groups with six rats in each group and were subjected to dermal treatment regimes for 30 days. Group I animals served as control and were applied with distilled water whereas group II received metalaxyl @ 350mg/kg b.wt. (1/10 LD₅₀) dermally after dissolution in water.

Enzyme assay

The rats were anaesthetized with diethyl ether and about 4-5 ml blood were collected from retro-orbital fossa and heart of anaesthetized rats in dry set of test tubes containing heparin @ 5-10 IU/ml of blood on 30th day of treatment. The plasma was immediately separated by centrifugation at 3000 rpm for 15 min and remaining red blood cells were washed with normal saline solution three times, before preparing the RBC lysate. Washing of erythrocytes were undertaken by diluting RBC sediment with normal saline solution in the ratio of 1:1 on v/v basis and centrifuged for 10 minutes after gentle but through mixing. After centrifugation the supernatant was discarded along with buffy coat and again NSS was added to the RBC on v/v basis, mixed gently and centrifuged. This process was repeated for 2-3 times. After final washing 1 per cent hemolysate (100µl washed RBC + 9.9 ml PBS) and 33 per cent hemolysate (330µl washed RBC+ 670µl PBS) in phosphate buffer solution (PBS), pH 7.4 were prepared. The 1 per cent haemolysate was used for the estimation of catalase, superoxide-dismutase, glutathione-peroxidase and glutathione-S-transferase and 33 per cent haemolysate was used for estimation of lipid peroxidation. The activity of lipid peroxidation in erythrocytes was determined according to method described by Shafiq-ur-Rehman (1984). The activity of superoxide dismutase (SOD), catalase (CAT), glutathione peroxidase (GPx) and glutathione-S-transferase (GST) in erythrocyte lysate were determined by methods of Marklund and Marklund (1974), Aebi (1983), Hafeman *et al.* (1974) and Habig *et al.* (1974) respectively.

Statistical analysis

The difference between two means based on individual observations was determined by unpaired Student's t-test. The significance was assayed at P < 0.05 and P < 0.01 levels (Snedecor and Cochran, 1967).

RESULTS AND DISCUSSION

Oxidative stress parameters were evaluated and the data obtained have been given in table 1. Perusal of the table revealed significant (P<0.01) increase in MDA value in metalaxyl treated animals as compared to control after 30 days of dermal application. Blood glutathione decreased significantly (P<0.01) in metalaxyl treated animals as

compared to control. The activity of Catalase (CAT) decreased significantly (P<0.01) after 30th day of dermal treatment with metalaxyl. Similarly the activities of Superoxide Dismutase (SOD) and Glutathione Peroxidase (GPx) significantly (P<0.05) in treated animals as compared to control. No significant alteration was observed in concentration of Glutathione-S-transferase (GST) in metalaxyl treated animals as compared to control. Malondialdehyde (MDA) is the end point of lipid peroxidation process which may be defined as an oxidative deterioration of polyunsaturated lipids (Dar *et al.*, 2013). Glutathione (GSH) an important antioxidant, protecting the membrane from oxidative insult, is thus considered as a critical determinant for the threshold of tissue injury caused by environmental chemicals (Machlin and Bandlich, 1987). Several enzymatic antioxidant defences designed to scavenge reactive oxygen species (ROS) in the eukaryotic cells protect them from oxidative injury. A fine balance between several antioxidant species and ROS appears to be more important for the overall protection of cells. Repeated dermal application of metalaxyl in rats caused a significant increase in lipid peroxidation on 30th day of experimentation as compared to the control group of rats. These findings are in agreement with studies on metalaxyl by Lamfon (2011) and Hashem (2012). According to Calviello *et al.* (2006) fungicide-induced damage is closely associated with increase in lipid peroxidation and the decrease in the antioxidant enzymes.

Table 1. Effect of repeated dermal application of metalaxyl on oxidative stress parameters in rats

Parameters/Units	Control	Treatment
Lipid peroxidation (n mol MDA formed/ml erythrocytes)	7.12±0.35	13.14±1.27 ^b
Blood glutathione	75.14±5.36	50.84±4.78 ^b
SOD (Units/mg protein)	59.20±3.71	40.82±5.69 ^a
CAT (µmole H ₂ O ₂ decomposed /min/mg protein)	60.05±5.59	35.57±3.77 ^b
GPx (Units/mg protein)	18.82±2.02	11.83±1.33 ^a
GST (µmole of conjugate of GSH-CDNB/min /mg plasma protein)	0.064±0.012	0.054±0.011

Values given are mean ± SE of the results obtained from 6 animals unless otherwise stated. ^{a,b} significantly different as compared to control values at 5% (P<0.05) and 1% (P<0.01) level of significance respectively.

Glutathione is a tripeptide comprising of glutamic acid, glycine and cystine, found in all tissues and occurs in approximately 2 mM concentrations in red blood cells (Beutler, 1975). GSH is an important naturally occurring antioxidant, which prevents free radical damage and helps detoxification by conjugating with chemicals. In addition, GSH is pivotal to the cellular antioxidant defenses by acting as an essential cofactor for antioxidant enzymes including glutathione peroxidase (GPx) and glutathione-s-transferase (GST) (Mascio *et al.*, 1991 and Hayes *et al.*, 2005). Under oxidative stress, GSH is depleted by GSH related enzymes to detoxify the peroxides produced due to increased lipid peroxidation (Cathcart, 1985). Decreased blood glutathione level has been observed due to metalaxyl oral treatment in rats (Hashem, 2012). A significant decrease in glutathione levels in rats exposed to benomyl has also been observed by Banks and Soliman (1997). SOD levels in metalaxyl-treated group of rats decreased significantly after after 30th day of exposure as compared to their control group. These findings are in consonance with the studies of Lamfon (2011) and Saber *et al.*

(2011) in metalaxyl-treated albino mice. Sakr and Abel-Samie (2008) has also found that mancozeb fungicides induce a significant decrease in the serum antioxidant superoxide dismutase. Catalase is a haeme-containing enzyme that catalyzes the dismutation of hydrogen peroxide into water and oxygen. The enzyme is found in all aerobic eukaryotes and is important for the removal of hydrogen peroxide generated in peroxisomes (microbodies) by oxidases, involved in β -oxidation of fatty acids, the glyoxylate cycle (photo-respiration) and purine catabolism. Stress conditions in which there is a large free radical generation also result in the depletion in catalase activity (Hertwig and Feirabend, 1992), thus justifying the decreased activity of this enzyme. Glutathione peroxidase is a selenium containing enzyme which reduces hydrogen peroxide forming GSSG and thereby serves as an alternative means of detoxifying activated oxygen. The activity of GPx is dependent upon glutathione level. Decreased glutathione activity in present study might be the reason for decreased activity of GPx. Repeated oral administration of metalaxyl in rats caused non significant decrease in GST levels on 30th day of experimentation. However findings of Hashem (2012), Calviello *et al.* (2006) and Sakr and Abel-samie (2008) revealed significant decrease in GST after oral treatment with metalaxyl.

Acknowledgement

The authors wish to express their deep sense of gratitude to the Vice chancellor and Dean, Faculty of Veterinary Sciences, SKUAST-Jammu for providing necessary facilities and granting permission to take up this work.

REFERENCES

- Aebi, H.E. (ed.). 1983. *Catalase. In: Methods of Enzymatic analysis*, pp. 276-86. Academic Press, New York.
- Banks, D. and Soliman, M.R.I. 1997. Protective effects of antioxidants against benomyl-induced lipid peroxidation and glutathione depletion in rats. *Toxicology*, 116(1-3): 177-181
- Beutler, E. 1975. Red cell metabolism. *A Manual of Biochemical Methods*. pp. 67-69. Grune Strotan, New York.
- Calviello, G., Piccioni, E., Boninsegan, A., Tedesco, B., Maggiano, N., Serini, S., Wolf, F.I. and Palozza, P. 2006. DNA damage and apoptosis induction by the pesticide mancozeb in rat cells: involvement of oxidative mechanism. *Carcinogenesis*, 28(6): 1202-1209.
- Cathcart, R. F. 1985. Vitamin C: the nontoxic, non rate-limited, antioxidant free radical scavenger. *Medical Hypothesis*, 18: 61-77.
- Dar, M.A., Khan, A.M., Raina, R., Kumar, P and Sultana, M. 2013. Effect of repeated oral administration of bifenthrin on lipid peroxidation and antioxidant parameters in wistar rats. *Bulletin of Environmental Contamination and Toxicology*. 90 (5). Online
- F.A.O./W.H.O. 1995. Pesticide residues in food — 1995. Report of the joint meeting of the FAO panel of experts on pesticide residues in food and the environment and the WHO core assessment group on pesticide residues, Rome, Italy, 11-20 September 1995.
- Habig, W.H., Pabst, M. J. and Jakoby, W.B. 1974. Glutathione-S-Transferase: The first enzymatic step in mercapuric acid formation. *Journal of Biology and Chemistry*, 249: 7130-7139.
- Hashem, H.E. 2012. Light and electron microscopic study of the possible protective effect of nigella sativa on metalaxyl induced hepatotoxicity in adult albino rats. *Journal of Cell Science and Therapy*, 3(2): 1-6.
- Hafeman, D.G., Sunde, R.A. and Hoekstra, W.G 1974. Effect of dietary selenium on erythrocyte and glutathione peroxidase in rats. *Journal of Nutrition*. 104: 580-587.
- Hayes, J. D. and Paiford, D. J. 1995. The glutathione-S-transferase supergene family. Regulation of GST and the contribution of the isoenzymes to cancer chemoprotection and drug resistance. *Critical Reviews in Biochemistry and Molecular Biology*, 30: 445-600.
- Hertwig, B. and Feirabend, J. 1992. Light dependence of catalase synthesis and degradation in leaves and the influence of interfering stress conditions. *Plant Physiology*, 100: 1547-1553.
- Howard, P.H. (ed.). 1991. *Handbook of Environmental Fate and Exposure Data for Organic Chemicals*, pp. 10-11. Lewis Publishers, Chelsea, MI.
- Lamfon, H.A. 2011. Protective effect of ginger (*Zingiber officinale*) against metalaxyl induced hepatotoxicity in albino mice. *Journal of American Science*, 7(6): 1093-1100.
- Machlin L and Bandlich A. 1987. Free radical tissue damage. Protective role of antioxidant nutrients. *Faseb J* 1: 441-445.
- Marklund, S. and Marklund, M. 1974. Involvement of superoxide anion radicle in autoxidation of pyrogallol and a convenient assay of superoxide dismutase. *European Journal of Biochemistry*, 47: 469-474.
- Mascio, P., Murphy, M. E., Sies, H.1991. Antioxidant defense systems: the role of carotinoids, tocopherols, and thiols. *American Journal of Clinical Nutrition*, 53:194-200.
- Okdah, Y.A. 2005. Effect of antox on metalaxyl fungicide induced histological and histochemical changes in liver of albino mice. *Journal of the Egyptian German Society of Zoology*, 48: 205-216.
- Punareewattana, K, Smith, B. J., Blaylock, B. L., Longstreth, J., Snodgrass, H. L., Gogal, R. M., Prater, R. M. and Holladay, S. D. 2001. Topical permethrin exposure inhibits antibody production and macrophage function in C57B1/6N mice. *Food and Chemical Toxicology*. 39: 133-139.
- Saber, S.A., Hawazen, L.A. and Amina, E.E. 2011. Ginger (*Zingiber officinale*) extract ameliorates metalaxyl fungicide induced nephrotoxicity in albino mice. *African Journal of Pharmacy and Pharmacology*, 5(2): 104-112
- Sakr, S.A. and Abel-Samie, H.A. 2008. Apoptosis related protein Bax in liver of metalaxyl fungicide treated mice: the effect of antox. *Ozean Journal of Applied Science*, 1(1): 17-27.
- Science News. 1994. Another emasculating pesticide found. 6:16.
- Shafiq-ur-Rehman. 1994. Lead induced regional lipid peroxidation in brain. *Toxicology Letters*, 21(3): 333-337.
- Sukul, P. and Spitteller, M. 2000. Metalaxyl: persistence, degradation, metabolism and analytical methods. *Reviews of Environmental and Contamination Toxicology*, 164:1-26.
- Snedecor, G. W. and Cochran, W. G. 1967. *Statistical Methods*. 6th ed. Ames: Iowa State University Press.
- U.S. National Library of Medicine. 1995. Hazardous Substances Databank, pp 9-10. Bethesda, M.D.